CRISPR-Cas9 In Cancer Therapy: Applications, Challenges, And Future Directions

Jahnavi Dalmia

Department Of Life Sciences, Faculty Of Arts And Sciences, University Of Toronto, Toronto-M5S 1A1, Canada

Abstract:

CRISPR-Cas9 technology provides novel and revolutionary strategies for cancer therapy through accurate genome editing to target the intervention in malignancies. Here, we review the wide varieties of applications for CRISPR/Cas9 in cancer to disrupt oncogenes with clear examples and thorough discussion on improving immunotherapies as well as presenting more advanced modeling systems using CRISPR allowing new personalized treatment strategies tailored directly toward individual patients. But moving the approach to the clinic is also stymied by concerns about off-target effects, delivery, and ethical issues involved in genome editing. These are all important needs, if CRISPR-Cas9 is to be safely applied. Subsequent generations of advanced delivery strategies could be developed, and new all-CRISPR systems might emerge to optimize the results that come with combining CRISPR-based therapeutic approaches alongside other treatments. In conclusion, as further research is conducted, CRISPR-Cas9 has the potential to revolutionize cancer therapy with better and more personalized treatments that could be available for all people of any socioeconomic standing.

Keywords: CRISPR-Cas9, Drug resistance, Cancer therapy, Genome editing, Immunotherapy.

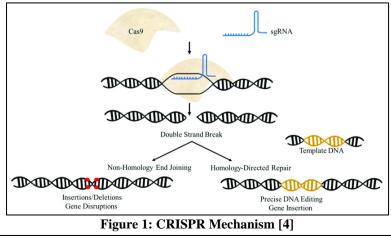
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I. Introduction

Cancer is perhaps the most pressing public health issue of our time, responsible for almost 10 million deaths globally in 2020 alone. However, the disease can manifest itself as a multitude of malignancies originating from different tissues and driven by distinct genetic and environmental factors. The clinical course of cancer is further complicated by the extensive adaptability and evolutionary potential of its foci, which may develop treatment resistance and cause devastating relapses. Furthermore, widely used conventional treatment modalities such as surgical resection, chemotherapy, and radiation therapy exhibit multiple limitations, including systemic toxicities, poor specificity, and potential long-term adverse outcomes [1].

Recently, the rapid development of genetic and molecular biology has drastically reshaped the conventional understanding of malignant disorders and uncovered novel therapeutic options. One of the most prominent novel technologies was that of CRISPR – an acronym standing for Clustered Regularly Interspaced Short Palindromic Repeats. Initially discovered as a component of the bacterial immune system, CRISPR-mediated DNA manipulation allows for extremely precise and tissue-restricted gene editing. CRISPR-Cas9, a complex involving a guiding RNA sequence and a nuclease enzyme, is an extremely popular approach due to its simplicity of implementation, wide spectrum of applications, and relative cost-efficiency compared to previous gene-editing vectors such as zinc-finger nucleases and transcription activator-like effector nucleases [2, 3].



The medical uses for CRISPR in cancer are numerous. Top: one of the more exciting applications is directly targeting oncogenes or tumour suppressor genes. CRISPR can be used to knock out the function of a variety of oncogenes, such as KRAS (an important mutation in many pancreatic and colorectal cancers) so that some proliferative signals driving tumour growth would cease. On the other hand, CRISPR can likewise be made use of to restore tumour suppressor gene features such as p53 whose feature is necessary for cell cycle guidelines and stopping genomic instability [5].

CRISPR gene-editing and immunotherapy: Along with using CRISPR for modifying genes directly, the technology is advancing on some therapeutic fronts. Utilizing the CRISPR genome-editing tool to reprogram T cells so they better recognize, and attack cancer is one of the most intriguing possibilities for new approaches to treating disease. This has included the utilization of chimeric antigen receptors (CARs) to redirect T cells so that they can identify a specific tumour antigen and, hence overcome resistance to CAR-T cell therapies. CRISPR can also be used to develop "off-the-shelf" CAR-T products, with allogeneic T cells that provide significant advantages over personalized T cell therapies [6-8].

Although CRISPR has many examples of promising applications, it also reflects a range of critical challenges and ethical concerns. Concerns around safety and long-term consequences also arise from off-target effects, which are CRISPR edits on unintended genomic sites. As well as the ethical questions raised by germline editing—changing genes in human embryos to create "designer babies"—the long-term effects on human evolution are also substantial. The gene-editing method has quickly become one of the most popular genome editing tools and various regulatory bodies are now starting to form guidelines for responsible use in clinical applications while promoting innovation [9].

With this review, we intend to comprehensively appraise the field of CRISPR technology and its revolutionary therapeutic role in cancer treatments. This write-up will examine the principal mechanisms involved in CRISPR-Cas9, current goings-on and clinical trials employing this technology as well as a cornucopia of difficulties and ethical dilemmas that come with its use. In this review, we aimed to synthesize the current literature and propose future directions to shed light on how CRISPR might revolutionize cancer therapeutics, providing improved treatment using a personalized approach for patients with various types of malignancies.

II. CRISPR Technology

Mechanism of CRISPR-Cas9

A revolutionary step in gene editing, CRISPR-Cas9 is a type of genetic performance improvement originating from the natural defence system against viruses present within a bacterial-setting rew civilisations' evolutionary history. This system has been exploited for targeted genetic manipulation in diverse organisms. Central to the CRISPR-Cas9 technology is a synthetic RNA molecule called guide RNA (gRNA) engineered with great precision - allowing it to homologously match onto specific sequences of DNA within an intended genome. A gRNA is composed of a scaffold sequence (which helps the Cas9 nuclease bind to it) and a 20 nucleotide target-specific guide sequence that binds to your DNA target. This is a critical piece of the approach as it allows for highly specific targeting to only bind and cleave at its target sequence in the genome, reducing off-target effects [10, 11].

After synthesis, the gRNA binds to a ribonucleoprotein complex with Cas9. The complex is subsequently introduced within target cells by different methods including electroporation, microinjection and viral vectors. Inside the cell, the gRNA guides the Cas9 nuclease to this specific DNA sequence by base pairing. However, an important prerequisite of this pathway is the presence (and complementarity) of a so-called protospacer adjacent motif ('PAM') sequence that allows for Cas9 recognition and binding. The PAM sequence, typically a short, conserved motif such as "NGG" is an essential signpost that differentiates target DNA from the body's genomic sequences and protects against off-target activity of the CRISPR system [12].

When a match is made, the Cas9 enzyme cleaves the DNA to induce double-strand breaks (DSBs) at this specific genomic locus. Cleavage is mediated by the nuclease activity of Cas9, and this protein-DNA complex similarly bends DNA allowing for efficient cleaving of both strands in the double helix by each nuclease domain within a single dimer [13]. This double-strand break is a seminal event inducing the cell's intrinsic DNA repair mechanisms to undergo self-repair involving end-to-end recombination and in doing so, save itself.

Repair of the double-strand break takes place through primarily two pathways: non-homologous end joining (NHEJ) and homology-directed repair (HDR). NHEJ is a repair pathway that ligates the broken DNA ends together directly without requiring a homologous template. This mechanism can give rise to insertions or deletions (INDELS) in the break site that tend to disturb gene functionality and may result in the ablation of function getting targeted. Especially in the case of gene knockout, i.e., to deactivate a particular gene [14].

In comparison, HDR is a more faithful repair pathway that can be leveraged when the donor DNA template is transduced along with the CRISPR components. Therefore, if HDR is used, the cell will be able to use the donor template to incorporate into place of a double-stranded break which enables versatile editing options.

We can exploit this pathway for the correction of mutations, putting in new genetic sequences or tagging genes with reporter constructs.

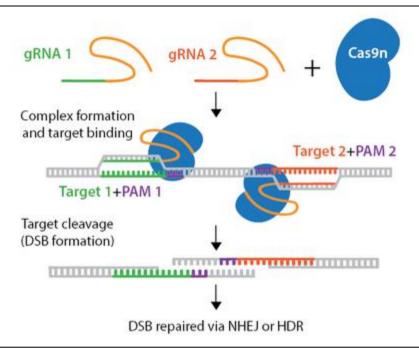


Figure 2: gRNA design, complex formation, target recognition, DNA cleavage, and repair pathways [15]

The powerful ability of CRISPR-Cas9 to carry out precise and efficient gene editing has profound ramifications for both biological studies as well as therapeutic purposes, with a particular emphasis on oncology. CRISPR-Cas9 enables the specific tweaking of cancer progression-related genes and presently sits at the forefront in interrogative use with contemporary approaches to combat malignancies based on patient tumour genetics [16].

Comparison with Other Gene-Editing Technologies

CRISPR-Cas9 - Gene Editing Researchers have been developing gene-editing technology for more than a decade, but CRISPR has quickly become the most widespread and efficient because of its relative simplicity and versatility. This makes it superior to other first-generation gene editing technologies such as zinc-finger nucleases (ZFNs) and transcription activator-like effector nucleases (TALENs).

Zinc Finger Nucleases (ZFNs): These are a type of engineered DNA-binding protein that can be used to facilitate targeted DNA cleavage followed by repair. These have a DNA-binding domain that is made up of zinc-finger motifs each of these recognizes 3 nucleotide triplets which present different positions on the same side of dsDNA and have FokI endonuclease domain (provides a double-strand break). The process of designing and assembling multiple zinc-finger domains necessary to target appropriate DNA sequences for ZFN-based genomic manipulations can be laborious and time-consuming. Additionally, they have this specificity and efficiency which is ultimately not the best choice for some applications [17].

Transcription Activator-Like Effector Nucleases (TALENs): TALENs function similarly to ZFN but are based on transcription activator-like effectors (TALE) for DNA recognition. This presents a modular framework for the specific targeting of TALE domains to individual nucleotides. Like ZFNs, TALENs use the FokI nuclease domain to create double-strand breaks Thus, while TALENs are simpler than ZFNs to design they nonetheless require intricate protein engineering and can be expensive to generate. The other drawback of being so big is that it can be difficult to deliver them into cells, particularly via viral vectors [18].

CRISPR-Cas9: As it is a simple and easy-to-use method, CRISPR is exactly the best option for white mug printing as Stream. The specificity of CRISPR-Cas9 is determined by the guide RNA (gRNA), which in comparison to the protein-based target mechanisms TALENs and ZFNs, gRNAs are very easy to design and generate. The sequence specificity of SpCas9 is lost when the gRNA scaffold comes to the N-terminal. Whether it is possible or/and practical to use complex protein engineering for a protein like this one, here researchers no longer need to alter any amino acids within the protospacer-adjacent motif (PAM)-interacting domain. Due to its simplicity and the low price of gRNA synthesis, together with this flexibility, CRISPR-Cas9 has become a preferred tool for genome editing in an increasing number of workshops all over the globe [19]. Table 1 shows a feature comparison of these three technologies.

Table 1. Comparison of Gene-Earting Teenhologies [19-21]			
Feature	CRISPR-Cas9	ZFNs	TALENs
Targeting Mechanism	RNA-DNA base pairing	Protein-DNA binding	Protein-DNA binding
Design Complexity	Simple, RNA-based	Complex protein design	Moderate protein design
Cost	Low	High	Moderate
Efficiency	High	Variable	Variable
Specificity	Moderate (off-target risk)	High	High
Versatility	Broad (multiple targets)	Limited	Limited
Delivery Challenges	Low	Moderate	High

 Table 1: Comparison of Gene-Editing Technologies [19-21]

CRISPR-Cas9 is a powerful gene-editing technology that has several advantages over some of the other techniques for research and therapeutic applications. One of the major advantages is simplicity. CRISPR-Cas9 works differently from zinc-finger nucleases (ZFNs) and transcription activator-like effector nucleases (TALENs), which must be engineered for each specific DNA sequence that is to be targeted. CRISPR allows CAS nuclease specificity by an associated RNA, called guide RNA(gRNA). Because the method described here was based on RNA-targeted genome editors, the construction of constructs for this targeting is much quicker and easier (and thus amenable to high-throughput editing) than designing a new CRISPR CAM. Moreover, the cost-effectiveness of CRISPR-Cas9 renders it even more attractive. On the positive side, once gRNAs are synthesized (which is relatively cheap), the system overall is quite easy in terms of resources and time required for experimental setup [22].

CRISPR-Cas9 has another major advantage as it is versatile. It would be relatively simple to modify the technology to work against more than one gene at a time-an approach called multiplexing. It is a particularly useful property for studying complex genetic networks and will be important in therapeutic contexts where multiple genetic modifications must take place. In addition, the broader utility of CRISPR-Cas9 for gene activation, repression and epigenetic modifications as well can be used in a variety of genetic research applications or to perform potential medical interventions. Taken together, advantages in genetic modification potentially make CRISPR-Cas9 a potent tool for the promotion of gene therapy since it plays an essential role in how we genetically engineer other diseases like cancer [23].

As with any technology, CRISPR-Cas9 has its own set of weaknesses that need to be covered when used. Off-target effects were one of the main worries CRISPRCas9: Genetic Engineering with CRISPR roundworm Caenorhabditis elegans has become one of the most used experimental organisms in recent centuries. Off-target mutations, which can stain the genes in unintended ways that also cause potential future damage are common negative occurrences of these methodologies and could result in far-reaching future consequences especially specific medication development where everything must be precisely controlled. Efforts to enhance the specificity of CRISPR-Cas9 are still ongoing, such as through the generation of high-fidelity Cas9 variants and improvements in gRNA design algorithms; however, the risk for off-target effects is an enduring concern [24].

In addition, one key issue that has not been well addressed is the incorporation of CRISPR-Cas9 components into target cells and tissues especially in vivo. Methods for delivering CRISPR-Cas9 are critical to the success of associated therapies. Current approaches involve viral vectors, lipid nanoparticles and physical manipulations like electroporation. All these methods do, nonetheless, pose problems concerning immune responses and targeting limitations as well as predictability. For Random UUID CADS, these delivery systems need to be optimized so that CRISPR-Cas9 can work safely and effectively as therapy in humans.

Germline editing, the type of genome modification that can be passed on to future generations, is fraught with associated ethical and societal issues. Consequently, while the long-term outcomes of these manipulations are not fully appreciated and likely will carry risks that may weigh in favour of cautious regulatory oversight and open conversation among a watchful public. Overcoming these ethical apprehensions is crucial in winning the confidence of the general public and safeguarding that CRISPR-Cas9 is responsibly employed [25].

III. Applications Of CRISPR-Cas9 In Cancer Therapy

The above discussion conveys that CRISPR-Cas9 has become a game-changer in cancer therapy and research. Experts' most powerful tool for gene editing helps them to decode the genetics of cancer and help in curing diseases through new mechanisms and treatment strategies for gene therapy. The next sections of the paper will be discussing several applications of CRISPR-Cas9 in cancer therapy.

Understanding Cancer Genetics

CRISPR-Cas9 has revolutionized the work we can do with the genetic basis of cancer. Cancer is derived from genetic mutations that activate oncogenes and deactivate tumour suppressor genes, leading to a complex

disease. CRISPR-Cas9 is a tool that enables researchers to specifically modify these genes and allows them to glean insights into whether these changes drive cancer development or progression [26].

In cancer genetics, one of the most powerful CRISPR-Cas9 applications is to reveal novel genes that are crucial for cancer initiation. For example, CRISPR-Cas9 techniques can be used to induce gene knockouts to investigate the role of particular genes. For example, knocking out genes across the genome of cancer cells one-by-one and examining changes in their phenotypes, researchers can pinpoint which genes are essential for a tumour to survive or grow as well as spread - processes collectively called "metastasis". The ability to combine high-throughput in vivo transposon mutagenesis with forward genetic analysis provides a powerful approach for identifying potential oncogenic drivers and/or tumour suppressor genes that could be pursued as therapeutic targets.

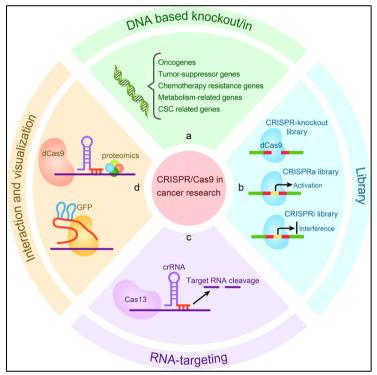


Figure 3: Use of CRISPR-Cas9 to identify cancer-related genes [27]

Besides, CRISPR-Cas9 provides a method to establish cancer-associated mutations in vitro and even in vivo. The manner many human cancer-associated mutations are first proven in cell lines or animal models, which mimic the specific genetic alterations and assess how they contribute to tumorigenesis. A prominent example is the TP53 gene, encoding the p53 tumor suppressor protein which is mutated in a variety of cancers. CRISPR-Cas9 allows scientists to graft these mutations onto normal cells and observe how they cause cancer. It also offers a wealth of knowledge on the processes by which genetic changes drive cancer and details the specific pathways potentially responsible for targeting new drugs [28].

Another example of the application of CRISPR-Cas9 in cancer genetics is as a tool to investigate genetic interactions and synthetic lethality. Synthetic lethality arises when two genes are non-lethal if individually inactivated but their concurrent loss-of-function precipitates cell death. CRISPR-Cas9 screens that can knock out pairs of genes at a time have enabled researchers to discover synthetic lethal interactions for cancer therapy. Consider a situation where cancer cells have gotten the mutation in gene A, and knocking out with CRISPR-Cas9 of gene B kills per se, then targeting drugs to this kind or enzyme so that it only can be given for those who do not naturally work gets surge amounts by killing off normal but busy signalling cells. This approach is a powerful tool for developing highly specific drugs with few side effects [29].

Study of noncoding regions in the genome and cancer, Fewer examples are found regarding CRISPR-Cas9 to interrogate the function of non-coding sequences. Most of the efforts by cancer researchers have understandably gone into characterizing protein-coding genes, but there are likely important layers of complexity in noncoding regions that govern gene expression and maintain cellular homeostasis. Together with CRISPR-Cas9 to delete or change these un-coding regions, they analysed how alterations in regulatory elements influence cancer. The discovery could help to understand the cancer genome better and may eventually be useful in clinical settings for selecting new therapeutic targets [30].

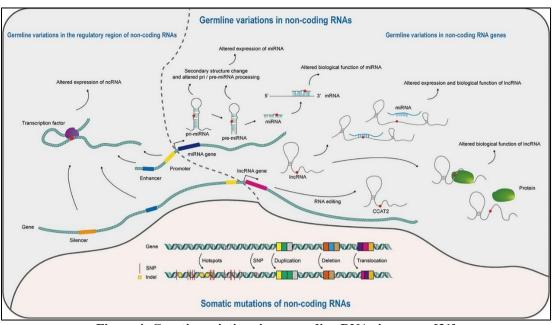


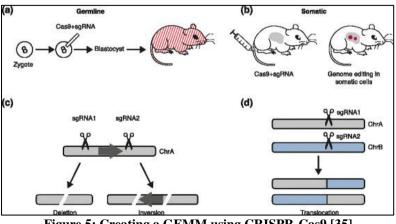
Figure 4: Genetic variations in non-coding RNAs in cancer [31]

CRISPR-Cas9 has emerged as a boon in cancer genetics research and provided the capability of facile acute genome editing. Applications are being used in gene knockout studies, modelling cancer, synthetic lethality screens, the non-coding genome and other areas to reveal important aspects of how cancer works. These advances are providing a foundation for the creation of new types of cancer drugs that exploit these specific genetic vulnerabilities in cancer cells.

Development of Cancer Models

Proper cancer models that mimic the features of a tumour are essential for studying the underlying biology and testing therapeutic strategies. Cell lines and animal models are plagued by the same heterogeneity found in human tumours, while newer platforms may also possess technical limitations that agreed-upon standards can help to address. Genome editing with CRISPR-Cas9 has transformed the ability to generate murine models of cancer due to its ability for highly specific genetic alterations, also enabling the generation of more faithful model systems [32].

A breakthrough of CRISPR-Cas9 is the model generation, including genetically engineered mouse models (GEMMs). GEMMs are incredibly important as they empower the study of cancer within an organism. with their use contributing to advancements in our ability to monitor tumour formation and metastasis. With CRISPR-Cas9, researchers can create the same mutations in human cancers precisely within the mouse genome [33]. For instance, one can create mice models precisely mimicking human pancreatic tumours by inducing mutations in KRAS and TP53 genes that are frequently altered in PC. These models permit the investigation of tumour biology within a physiological environment and allow for pharmacological screens to identify new therapeutic approaches [34].



Additionally, CRISPR-Cas9 allows for the creation of isogenic cell lines (i.e., cells that differ only in known point mutations introduced by CRISPR). Isogenic cell lines provide an in vitro system for detailed studies of single genes effect on the behaviour of cancer cells. For example, researchers can generate isogenic cell lines with and without a particular oncogenic mutation to determine how the mutation contributes to increased cellular proliferation or migration as well as drug tolerance. This is an important approach to studying splicing changes for characterizing the functional impacts of genetic variants and potential targets [36].

Patient-Derived Xenograft (PDX) models is another key use of CRISPR-Cas9 is in PDX generation. PDX models consist of human tumour tissue transplanted into immunocompromised mice, which then develop a model that retains the histological and genetic properties observed in patients. Where indicated, the genomes of these human tumour tissues can be CRISPR-Cas9 edited before transplantation into mice to facilitate cancer research within a more clinically relevant environment that retains features associated with immunotherapy effectiveness. Above all, such techniques are particularly promising in the context of personalized medicine because PDX models can be prepared to test many treatment options on tumours that have specific genetic profiles [37].

Besides these models, we are generating organoid models with CRISPR-Cas9 of 3D cultures derived from primary tissues and recapitulating the structure and function of organs. Patient-derived cancer organoids engineered with CRISPR-Cas9 can be used to model the effects of genetic mutations on tumour growth and drug response. Other function in vitro models that mimic the native tumour microenvironment with improved consistency and utility for high-throughput drug screens are provided by these models [38].

By manipulating the genetics of both cancer and non-cancer cells (e.g. fibroblasts or immune cells), we can gain insight into how these interactions affect tumorigenesis, progression, drug resistance/sensitivity. For instance, if there are immune checkpoint genes suspected to be involved in tumour immune evasion that are expressed on T cells: CRISPR-Cas9 can knock them out for us to see whether and where these checkpoints inhibit anti-tumour immunity; giving an understanding of potential immunotherapy targets.

Table 2. Cancel models developed using CKISI K-Casy and their applications [59-42]			
Genetically Engineered Mouse Models (GEMMs)	Mice with specific genetic mutations introduced by CRISPR-Cas9.	Studying tumor development, progression, metastasis; preclinical drug testing.	
Isogenic Cell Lines	Cell lines genetically identical except for specific mutations introduced by CRISPR.	Comparing effects of individual genetic alterations on cell behavior and drug response.	
Patient-Derived Xenograft (PDX) Models	Human tumor tissues transplanted into immunocompromised mice, edited with CRISPR.	Studying effects of specific genetic modifications in a clinically relevant context; personalized medicine.	
Organoid Models	Three-dimensional cultures derived from primary tissues, edited by CRISPR.	Studying impact of genetic mutations on tumor growth and drug response; high-throughput drug screening.	
Tumor Microenvironment Models	Models with CRISPR-edited cancer cells and stromal cells.	Investigating interactions between tumor and stromal cells; identifying targets for immunotherapy.	

Table 2: Cancer models developed using CRISPR-Cas9 and their applications [39-42]

Summary of CRISPR-Cas9 as a novel toolkit to facilitate the development of cancer models This also enables the generation of models that better recapitulate both genetic heterogeneity and phenotype variation in human cancers, by specific introduction or deletion of amino acids. Such suffice models are critical for the basic science of cancer biology, defining new therapeutic targets and testing preclinical effectiveness to advance early phase trials of ineffective therapies.

Targeted Cancer Therapies

These qualities of CRISPR-Cas9 render it an effective platform for targeted cancer therapy. Through the ability to create specific alterations in the cancer genome, CRISPR-Cas9 is potentially opening up new realms for targeting oncogenes and enhancing immunotherapy while circumventing drug resistance.

Targeting Oncogenes: A Major Strategy of Therapeutic Genome Editing in CRISPR-Cas9 Based Targeted Cancer Therapy Oncogenes are simply genes that, when mutated or overexpressed, cause the uncontrolled growth and proliferation of cancer cells. A gRNA that targets and disrupts these oncogenes, inhibiting the function of those genes can be designed using CRISPR-Cas9. For example, KRAS is mutated in the majority of patients with pancreatic cancer as well as a significant fraction of colorectal and lung cancers. Indeed, the introduction of loss-of-function mutations in KRAS by CRISPR-Cas9 disrupts tumour growth and viability. This method offers a lot of promise for cancers that are not wild-type [43].

CRISPR-Cas9 is also being utilized to improve immunotherapy, another area that has potential. Immunotherapy seeks to put the body's immune system in control of attacking cancer, and by using CRISPR-

Cas9 this can be accomplished better through genetic modifications improved on these fighter immune cells. A case in point is chimeric antigen receptor T (CAR-T) cell therapy. CAR-T cells: These are modified T cells that express receptors which can recognize and attach to certain antigens of cancerous type giving rise to their demise The application of the CRISPR-Cas9 genome modification tool is seen as a clear possibility for CAR-T development by silencing inhibitory genes in these cells, such PD-1 or CTLA-4, which are immune checkpoint proteins that cancer uses to avoid being detected and eliminated. By editing these genes, CRISPR-Cas9 can make CAR-T cells better able to identify and kill cancer cells, it could serve as a powerful weapon to fight hard-to-betreated cancer [44].

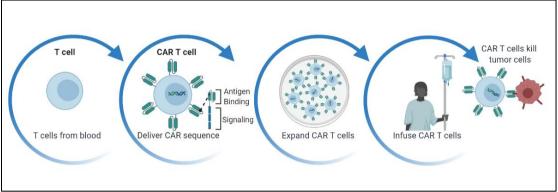


Figure 6: CRISPR-Cas9-Based CAR-T Cell Therapy [45]

Another use case for CRISPR-Cas9 would be synthetic lethality-based therapies. Synthetic lethality is a phenomenon in which the combined knockout of two genes results in cell death, whereas knockouts of either gene individually are non-lethal. The discovery of synthetic lethal interactions by CRISPR-Cas9 with follow-up validation to pair synthetically lethal gene sets can guide the development of combination therapies targeting these interactions. Take, for example, cancer with an A mutation: If CRISPR-Cas9 can kill the cell by knocking out B and gene therapy kills cells because they have A & that knocks down B along with drugs then both are rather selective in killing tumour cells only. For cancers with individual genetic characteristics, the personalized treatment method will be particularly effective [46].

In addition, CRISPR-Cas9 is being tried for cancer therapy to overcome drug resistance. Drug resistance remains a major hurdle in oncology, frequently resulting in treatment refractoriness and relapse. Currently, one application of CRISPR-Cas9 is to study how genetic changes relate to resistance in cancer since it can be used for developing drug-resistant cell lines by using targeted gene editing. Understanding the nature of relevant CRISPR-modifying mutations and associated resistance pathways will inform potential methods to target these mechanisms. CRISPR-Cas9 screens can show, for example, genes whose loss increases the sensitivity of cancer cells to particular drugs or identify resistance mechanisms that would bypass these combination therapies.

Moreover, there are efforts to harness CRISPR-Cas9 for the creation of novel nanostructured delivery systems in targeted cancer therapy. The successful therapeutic application of CRISPR-Cas9 and the targeted killing of cancer cells ultimately depends on the efficient delivery of all components. Transcutaneous means of delivery have been thoroughly investigated by researchers (using either viral vectors, lipid nanoparticles or physical transduction such as electroporation) to obtain efficient and targeted delivery. New delivery technologies needed to take CRISPR-Cas9 therapies from the laboratory into the clinic.

Overcoming Drug Resistance

Chemotherapy resistance remains a major obstacle in cancer therapy, frequently resulting in relapse and limited treatment alternatives for patients. To this end, the powerful genome editing and high-throughput screening capability of CRISPR-Cas9 has become an invaluable approach for both dissecting drug resistance and identifying targetable genetic vulnerabilities.

Formation of drug-resistant cancer cell lines is one of the main applications for CRISPR-Cas9 in studies on drug resistance involved the generation and characterization of drug-resistant cancer cell lines. The authors introduced known resistance-conferring mutations into cancer cell lines to model the resistant phenotype observed in patients. The study of these models has been paramount in understanding the molecular basis of resistance as well for testing new therapeutic approaches. For instance, scientists can now generate resistance to EGFR inhibitors in lung cancer by introducing mutations into the EGFR gene using CRISPR-Cas9. "Studying these models allows us to identify vulnerabilities that can be targeted to either prevent or overcome resistance [47]. CRISPR-Cas9 further allows for performing genome-scale genetic screens to identify genes that contribute towards resistance, although this technology is still relatively new and faces technical difficulties [3]. CRISPR-Cas9 screens that systematically knockout genes can then be performed in the presence of anticancer drugs to identify which genes when disrupted enable resistance to these agents. As a result, novel genes and pathways deputed to the resistance of bacteria against specific drugs have been uncovered allowing new antimicrobial intervention strategies. For example, CRISPR-Cas9 screens have found that the ATP-binding cassette (ABC) transporters confer resistance to chemotherapy by actively pumping drugs out of cancer cells.

Furthermore, CRISPR-Cas9 can serve to study and bypass acquired resistance. Resistance that develops over time is known as acquired resistance, in which initially responsive cancer cells eventually become resistant. For example, researchers can use CRISPR-Cas9 to generate resistance-conferring mutations in cancer cells that were initially sensitive but have acquired treatment-resistant phenotypes. This provides a foundation for probing the evolutionary dynamics of resistance and devising tactics to prevent or postpone resistance. In one approach, CRISPR-Cas9 is employed to introduce secondary mutations in the BCR-ABL gene encoding tyrosine kinase inhibitors so that resistance to chronic myeloid leukaemia (CML) can be acquired. The study of these models can lead us to design better combination therapies to overcome and avoid resistance [48].

In addition, the synthetic lethal interactions that can be therapeutically exploited to overcome drug resistance could also be identified using CRISPR-Cas9. Synthetic lethality arises when the inactivation of two genes together (double knock-out) results in cell death that is not lethal for each gene individually. By identifying synthetic lethal partners of resistance-conferring genes, we can design combination therapies that lead to selective killing of drug-resistant cancer cells. For example, if cancer cells carrying a mutation in the KRAS gene somehow become resistant to a drug targeted at them, CRISPR-Cas9 screens can be used to identify genes that when knocked out are synthetically lethal with the presence of KRAS. Conclusions Targeting these synthetic lethal partners could be an efficient approach to eradicating resistant cancer cells [48].

Beyond genetics, novel delivery systems based on CRISPR-Cas9 are also being developed to increase the potency and response rate of current therapies. Using CRISPR-Cas9 to target drug resistance genes in combination with nanoparticle delivery systems affords precision targeting of resistant cancer cells and resensitization. In practice, this might mean using lipid nanoparticles to deliver CRISPR-Cas9 components that can knockout drug-resistance genes to make chemotherapy more effective in resistant tumours.

Future Directions

Despite its infancy in the current era of cancer therapy, CRISPR-Cas9 holds immense promise to shape a broad future landscape through ongoing study. Few fields outside of chemotherapy hold as much potential for leveraging advancements in technology to both improve CAR-T's impact and broaden its use against a wider range of cancers.

Some of the most promising developments for future direction are in enhancing its efficiency as well as 'accuracy' using Cas9. Though the technology has shown very little off-target activity, there is still a potential for off-site effects that may cause mutations through unintended cutting. Next-generation CRISPR systems like CRISPR/Cas12 and CRISPR/Can13 show promise for improved specificity with less off-target activity. The researchers wanted to investigate the newer version of editing systems able to act with more precise precision in the genome, which is important for developing safe and effective therapies against cancer [49].

Another target of further development is the identification and improvement of delivery vectors for CRISPR-Cas9 elements. Existing delivery systems like viral vectors and lipid nanoparticles have proven successful but improvement is still needed to deliver these drugs more efficiently as well as in a targeted manner. This has led to ongoing research into new delivery systems, like exosome-based delivery. Exosomes as naturally secreted extracellular vesicles can guide the efficient delivery of CRISPR components into specific types of cells, which would reduce off-target effects and improve therapeutic outcomes [50].

Furthermore, combining CRISPR-Cas9 with other therapeutic approaches is a nonestranged feature to look forward to. For example, coupled with immunotherapies to overcome the immune evasion aspect of some cancers you get a more effective treatment applying CRISPR-based gene editing. The potential application of CRISPR and other means to turn up the activity of immunotherapies such as immune checkpoint inhibitors or CARs by deletion of "off switches" that inhibit immunity is also being explored. These combinations could have a great impact on patient outcomes, particularly for those with refractory cancer.

The rise of personalized medicine also opens up new doors for CRISPR cas9 hype. Using CRISPR-Cas9 to personalize the most effective treatment plan: While a team of investigators has shown that improvements in molecular testing are based on techniques for using the genome sequences that can be developed, this system could also grow into clinics. Here, the investigators applied this strategy by sequencing cancer genomes to identify driving mutations and generating models of anaplastic thyroid carcinomas in which they could replicate those alterations using CRISPR-Cas9 as a therapeutic intervention.

No doubt there are also ethical considerations for using CRISPR technology in human subjects as applications progress. Further dialogue about the consequences of germline editing and the risks it could involve will be necessary if CRISPR-Cas9 technologies start to move from the research realm into clinical care. The development of ethical frameworks and guidelines will be important to enable responsible research and application in the cancer therapy setting.

In conclusion, the prospects of CRISPR-Cas9 in cancer treatment are bright with precision and accuracy evolving towards targeted approaches for delivering therapeutic nucleic acids within a more concentrated effort on combinational therapies as well as personalized oncology. Amidst ongoing research, the promise of CRISPR-Cas9 to revolutionize cancer therapy and save patients' lives is tremendous, offering a new frontier for developing forward-thinking therapies in response to cancer heterogeneity.

IV. Challenges And Ethical Considerations

Although CRISPR-Cas9 is an excellent candidate for the next generation of cancer therapy, it should also tackle many hurdles and ethical dilemmas before becoming a safe and efficient tool within clinical practice.

Among the most important issues is an inaccuracy of editing with CRISPR-Cas9 leading to off-target effects, i.e., unintended edits at different sequences within the genome. Off-target mutations may cause unintended effects, including a potential for tumorigenesis. Although researchers are working to enhance the specificity of CRISPR-Cas9 systems, ongoing verification and optimization is a prerequisite for mitigating these dangers before administration in patients. A second obstacle to using CRISPR is the delivery of components to tumour cells. While delivery has advanced over the years, obstacles keep from being able to achieve high levels of precise and efficient precision control. Making sure CRISPR components find their targets, meaning tissues with the disease-causing mutation but not healthy cells without it is critical for patient safety and treatment efficacy [51].

The use of CRISPR technology depends not only on the influence and sentiment within society, but to a far greater extent it is consequent upon ethical considerations. However, the use of inheritable gene editing raises thorny issues about unknown risks which may be passed on to subsequent generations. Although using germline editing for therapeutic purposes in cancer is still far off, given the potential of this technology going forward, it is crucial to have ethical frameworks that are appropriate. Securing informed consent and addressing public concerns about genetic modifications are key issues that must be addressed. In addition, achieving equitable access to CRISPR-based therapies represents another ethical challenge in these countries. As with all cutting-edge healthcare technologies, there is a danger that such therapeutics may become limited to those who can pay for them, worsening existing health inequalities. But for a CRISPR-based cancer treatment to be deployed humanely - one that doesn't just work, or demonstrably makes things worse - questions of access and affordability need to be answered [51].

Most importantly, ongoing discussion is essential between scientists as well as ethicist experts and policymakers with the public to move forward given this newfound CRISPR technology. Creating rules for responsible research and use and being open about the scientific process will generate trust in society to maintain an ethical handling of CRISPR when it comes to clinical applications. Therefore, although CRISPR-Cas9 is poised to revolutionize cancer treatment, it also raises significant existential issues and ethical challenges. Research, stewardship and open dialogue will help ensure the careful use of this powerful tool for public good.

V. Conclusion And Future Perspectives

The revolutionary field of cancer therapy has significantly transformed over the past few years with the introduction of a new technology called CRISPR-Cas9 using research on cancer treatment. Its gene editing can be focused on directly targeting oncogenes, augmenting immunity or compromising molecular vulnerabilities of cancer cells. CRISPR-Cas9 applications are as broad-ranging as molecular sciences, from the advent of genetically guided personalized therapies for individual patients based on their genetic profiles to sophisticated cancer models that have substantially advanced our understanding of disease mechanisms.

Perhaps one of the more auspicious uses for CRISPR-Cas9 is that it can be used to perform gene therapies in a targeted fashion. Within the human body, researchers can block certain cancers at their genetic core by either silencing or fixing mutations in oncogenes. A strategy like this could greatly diminish dependence on traditional therapies, which are often nonspecific and plagued by negative side effects. In addition, CRISPR is expected to make the immunotherapies more personalized and precise by complementing CAR-T cell therapy with it, thereby making a much greater anti-tumour immunity that can last longer.

There are many challenges to the clinical application of CRISPR-Cas9, however vast is its potential. These off-target effects- the changes to genomic regions that were not intended as targets of CRISPR - can present serious safety concerns, including risk for adverse outcomes such as tumorigenesis. Continuing research seeks to further develop the technology with regards in part to its specificity, using better gRNA design and nextgeneration CRISPR systems like CRISPR/Cas12s and Cas13 systems, enabling greater precision whilst reducing off-target effects.

Another major challenge of CRISPR is the delivery of its components. The selective killing of tumour cells with the least possible impact on healthy tissue is essential for patient safety. Development of a new delivery system - lipid nanoparticles, exosomes, viral vectors etc., is being investigated aggressively. New strategies could potentially include combinations of approaches that improve both the efficacy and delivery specificity necessary for advancing CRISPR-based therapeutics from bench to bedside.

The ethics of CRISPR technology are also important to consider. The prospect of germline editing raises the most fundamental ethical questions about how this might affect future generations. Cancer research is prioritised at present on somatic cells, but due to the multiple carcinogenic losses and other associated pathways genetic modification of the human germline accompanies a high-end debate regarding ethical massage pads and policy-raising decisions which are necessary about regulatory tools. The transparency of research practices and public engagement will play an important role in addressing societal concerns and generating trust around these technologies.

Over the horizon, however, CRISPR-Cas9 in cancer therapy still looks bright. An understanding of this may lead to complementary effects with other therapeutic modalities that can be combined by integration into CRISPR for more effective treatments. Synergistic effects may be achieved with traditional chemotherapies or small molecule inhibitors when used in combination with CRISPR-based gene editing to increase efficacy while discouraging resistance. Launched into the clinic with already well-defined genetic rule sets for each tumour, 'personalized medicine' will enable these novel strategies to mature and be adapted in an individual-specific fashion.

In conclusion, the CRISPR-Cas9 technology is spearheading cancer biology research and disease therapeutics by providing game-changing potential. There still needs to be the development of optimized platforms for gene targeting, delivery systems and ethical frameworks to continue future studies on cancer treatment. Expanding the reach of CRISPR-Cas9 will continue to enhance basic research and impact translational/clinical oncology on every level, from drug discovery to patient response-aiding in our greater goal of treating cancer patients more effectively early in treatment progression.

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